

 -1 correlation coefficient $+1$

4. Results

4.1. Nucleotide composition in proto-oncogenes/oncogenes

The overall nucleotide compositions in the complete coding sequences of eighty two human proto-oncogene/oncogenes were analyzed (Table 3). The mean value of A base (520.8) was observed at the highest, followed by C (517.6), G (502.2) and T (416.6) among all the coding sequences. The GC content for the selected coding sequences ranged from 38.1% to 71.0%, with a mean of 54.1% and a standard deviation of 0.086. The AT content for the selected coding sequences varied from 29.0% to 61.9%, with a mean of 45.9% and a standard deviation of 0.086. We calculated the GC content at different codon positions (Figure 4) and it was found that the GC content at each codon position variation among the genes.

It is a well known fact that the nucleotide at the third codon position varies considerably due to the wobble hypothesis which allows the cell to identify the codons that encode 61 different amino acid using less than 61 tRNA molecules.

In our analysis, the comparison of the nucleotide composition at third codon position (A_3, T_3, G_3, C_3) confirmed that the mean value of C_3 (187.9) was the highest followed by G_3 (178.7), T_3 (150.9) and A_3 (134.9). The mean value of $GC_3\%$ (ranged from 18.2% to 90.8%) and AT3% (ranged from 9.2% to 69.6%) were 54.9% and 45.1% respectively with an exact standard deviation of 0.20. The average value of $(GC₁+GC₂)$ % ranged from 39.8% to 62.9% with a mean of 50.8% and a standard deviation of 0.057 which could be presumed that G/C ending codons might be preferred in coding sequences of proto-oncogenes/oncogenes.

Table 3: Nucleotide composition analysis in the coding sequences of eighty-two proto-oncogenes/oncogenes

CDS	A	Τ	G	С	A_3	T_3	G3	C_3	AT	GC	GC ₁	GC ₂	GC ₃	AT ₃	GC ₁₂
N0.									℅	$\%$	%	$\%$	%	$\%$	℅
1	817	580	982	1014	161	165	377	428	41.2	58.8	56.6	48.7	71.2	28.8	52.7
\overline{c}	955	745	935	914	280	292	302	309	47.9	52.1	56.8	47.8	51.6	48.4	52.3
3	324	260	437	422	24	50	197	210	40.5	59.5	57.6	36.4	84.6	15.4	47
4	104	85	124	131	18	21	55	54	42.6	57.4	50.7	48	73.6	26.4	49.3
5	272	184	171	189	96	81	50	45	55.9	44.1	50.4	47.1	34.9	65.1	48.7
6	609	429	711	759	128	126	265	317	41.4	58.6	57.9	48.3	69.6	30.4	53.1
7	114	132	238	236	13	28	98	101	34.2	65.8	64.2	50.4	82.9	17.1	57.3
8	206	215	372	548	37	52	136	222	31.4	68.6	70.7	55	80.1	19.9	62.9
9	458	385	479	631	77	137	172	265	43.2	56.8	56.1	47.3	67.1	32.9	51.7
10	849	623	1178	1166	96	119	521	536	38.6	61.4	58.8	42.4	83.1	16.9	50.6
11	660	579	526	536	230	220	162	155	53.8	46.2	53.1	44.1	41.3	58.7	48.6
12	871	552	1024	997	115	99	463	471	41.3	58.7	58.5	36.1	81.4	18.6	47.3
13	845	743	627	734	280	335	173	195	53.8	46.2	55.3	45.7	37.4	62.6	50.5
14	146	132	176	224	45	43	59	79	41	59	65	50.9	61.1	38.9	58
15	196	149	267	276	25	14	129	128	38.9	61.1	60.5	36.1	86.8	13.2	48.3
16	192	175	252	251	25	44	118	103	42.2	57.8	61.7	35.5	76.2	23.8	48.6
17	162	162	267	288	27	37	114	115	36.9	63.1	64.8	46.4	78.2	21.8	55.6
18	170	104	239	303	29	24	86	133	33.6	66.4	63.6	55.1	80.5	19.5	59.4
19	605	615	583	543	174	287	180	141	52	48	62.4	40.5	41	59	51.5
20	316	269	334	365	65	84	125	154	45.6	54.4	51.9	46.3	65.2	34.8	49.1
21	149	87	150	124	44	35	57	34	46.3	53.7	63.5	44.1	53.5	46.5	53.8
22	186	121	135	122	51	53	46	38	54.4	45.6	48.9	43.1	44.7	55.3	46
23	455	223	271	179	128	97	96	55	60.1	39.9	44.4	35.1	40.2	59.8	39.8
24	313	259	326	320	59	82	110	155	47	53	50.7	43.1	65.3	34.7	46.9
25	349	324	235	310	102	117	94	93	55.3	44.7	46.6	41.6	46.1	53.9	44.1
26	759	719	1120	1170	140	196	429	491	39.2	60.8	63	46.1	73.2	26.8	54.5
27	309	276	402	468	63	83	152	187	40.2	59.8	64.7	44.7	69.9	30.1	54.7
28	361	275	316	407	80	90	139	144	46.8	53.2	56.5	40.6	62.5	37.5	48.6
29	972	748	707	729	263	318	240	231	54.5	45.5	50.1	41.6	44.8	55.2	45.9
30	503	345	588	532	155	191	134	176	43.1	56.9	62.5	61	47.3	52.7	61.7

Table 3 continued

M: mean; SD: standard deviation; GC₁₂: average of GC contents of first and second codon positions

4.2. Nucleotide composition in tumour suppressor genes

We analyzed the nucleotide composition of the coding sequences of sixty-three tumour suppressor genes (Table 4) which revealed that mean value of A (767.2) was the highest followed by G (655.2), C (654.4) and T (613.2) among all the coding sequences. The overall mean percentage of GC (ranged from 35.8% to 74.7%) and AT (ranged from 25.3% to 64.2%) contents was 50.7% and 49.3% respectively with an equal standard deviation of 0.09. The GC contents at different codon positions were calculated and it was observed that the GC content at each codon position variation among the genes (Figure 5).

The nucleotide composition at the third position of codon (A_3, T_3, G_3, C_3) showed that the mean value of T_3 (233.5) was the highest followed by C_3 (227.7), G_3 (220.8) and A₃ (214.6). The GC₃ values (ranged from 26.7%-94.7%, mean=54.4%, SD=0.17) was compared with that of AT_3 values (ranged from 5.3%-73.3%, mean=45.6%, SD=0.17) in the coding sequences of tumour suppressor genes.

The average percentage of GC contents of the first and second codon position $(GC₁₂)$ was found to range from 37.4% to 67.6% with a mean value of 48.9% and a standard deviation (SD) of 0.06. Therefore, from the overall nucleotide composition analysis, it could be concluded that GC–ending and AT–ending codons are almost equally distributed in the coding sequences of tumour suppressor genes.

Table 4: Nucleotide composition analysis in the coding sequences of sixty-three tumour suppressor genes

Table 4 continued

M: mean; SD: standard deviation; GC₁₂: average of GC contents of first and second codon positions

4.3. Nucleotide skewness among proto-oncogenes/oncogenes

The variation in base composition within each coding sequence was calculated from their differences in usage for the GC, AT, keto, amino, purine, and pyrimidine bases of the eighty-two proto-oncogenes/oncogenes (Table 5). The magnitude of skew values revealed that base composition bias is linked to transcription process (Fujimori et al. 2005, Touchon and Rocha 2008). In our analysis, forty-three protooncogenes/oncogenes out of eighty-two showed negative GC skew value and the remaining thirty-nine showed positive GC skew value (Figure 6). Positive GC skew indicates richness of G over C and negative GC skew reveals richness of C over G (Tillier and Collins 2000).

In case of AT3 skew, nearly all the selected proto-oncogenes/oncogenes showed positive value which indicates richness of A over T (Tillier and Collins 2000). Negative value of keto skew was observed as reported earlier by Zhang & Gerstein in human genome (Zhang and Gerstein 2003). The keto skew values were negative for forty-three proto-oncogenes/oncogenes. A wide variation of purine, pyrimidine and amino skews was observed which might affect the expression patterns of these genes in the cell.

Table 5: Skew values of eighty-two proto-oncogenes/oncogenes

Table 5 continued

4.4. Nucleotide skewness among tumour suppressor genes

The variation in base composition in each coding sequence of tumour suppressor gene was calculated from their differences in usage for the GC, AT, keto, amino, purine, and pyrimidine bases (Table 6). Skew value reveals that the base composition bias is linked to transcription process (Fujimori et al. 2005, Touchon and Rocha 2008). In our analysis, twenty-nine tumour suppressor genes out of sixty-three showed the negative GC skew value and the remaining twenty-four showed positive GC skew value (Figure 7).

Figure 7: GC skew values of sixty-three tumour suppressor genes Positive GC skew indicates the richness of G over C whereas negative GC skew reveals the richness of C over G for tumour suppressor genes (Tillier and Collins 2000). In case of AT3 skew, nearly all the selected tumour suppressor genes showed positive value which depicts the richness of A over T (Tillier and Collins 2000). Negative value of keto skew was observed as reported earlier by Zhang & Gerstein in human genome (Zhang and Gerstein 2003). The keto skew values were negative for twenty-three tumour suppressor genes out of sixty-three. A wide variation of purine, pyrimidine and amino skew was observed which might affect the expression patterns of these genes.

Table 6: Skew values of sixty-three tumour suppressor genes

Continued

4.5. Codon usage patterns in the coding sequences of protooncogenes/oncogenes

The correlation coefficient between codon usage and GC bias was analyzed using a heat map (Figure 8) in order to find out the relationship between the codon usage variation and GC constraints among the selected coding sequences of human proto-oncogenes/oncogenes.

Figure 8: Heat maps of correlation coefficient of codons with GC_{3s}. The color coding red represents the positive correlation, green as negative correlation. The black fields are stop codons (TAA, TAG, TGA) and non-degenerate codons (ATG, TGG) in the coding sequences of the 82 proto-oncogenes/oncogenes under study.

 In our analysis, nearly all the codons ending with G/C base showed a strong positive correlation (p<0.01) with GC bias and all the A/T–ending codons showed negative correlation with GC bias. But, five C–ending codons (AAC, ACC, AGC, TCC, GTC) as well as five G –ending codons (AAG, AGG, TTG, CAG, GGG) showed a negative correlation between codon usage and GC bias. These codons display non-linear upward usage profiles due to the effect of GC bias (Figure 9). The codon TTG encoding leucine amino acid exhibited significant negative correlation (p<0.01) with GC_{3s}.

Figure 9: Scatter plots of negatively correlated codons with GC3s. [A] Scatter plots of codon usage frequency for the asparagine codon AAC $\text{vs } GC_{3s}$. [B] Scatter plots of codon usage frequency for the lysine codon AAG vs GC_{3s}. [C] Scatter plots of codon usage frequency for the arginine codon AGG vs GC_{3s}. [D] Scatter plots of codon usage frequency for the leucine codon TTG vs GC_{3s}.

4.6. Codon usage patterns in the coding sequences of tumour suppressor genes

To investigate the relationship between the codon usage variation and GC constraints among the selected coding sequences of human tumour suppressor genes, we analyzed the Pearson's correlation coefficient between codon usage and GC bias using heat map (Figure 10).

In our analysis, nearly all G- and C- ending codons showed strong positive correlation with GC bias, and conversely for A- and T- ending codons. However, one C–ending codon AAC (asparagine) and three G-ending codons namely AAG (lysine), AGG (arginine) and TTG (leucine) showed negative correlation between codon usage and GC bias. These codons exhibit non-linear upward usage profiles as a function of GC bias (Figure 11). The codons AGG and TTG encoding amino acid arginine and leucine respectively displayed significant negative correlation (p<0.01) with GC_{3s}.

Figure 10: Heat maps of correlation coefficient of codons with GC_{3s}. The color coding red represents the positive correlation, green as negative correlation. The black fields are stop codons (TAA, TAG, TGA) and non-degenerate codons (ATG, TGG) in the coding sequences of the 63 tumour suppressor gene under study.

Figure 11: Scatter plots of negatively correlated codons with GC3s. [A] Scatter plots of codon usage frequency for the asparagine codon AAC $\text{vs } GC_{3s}$. [B] Scatter plots of codon usage frequency for the lysine codon AAG $\text{vs }GC_{3s}$. [C] Scatter plots of codon usage frequency for the arginine codon AGG vs GC_{3s}. [D] Scatter plots of codon usage frequency for the leucine codon TTG vs GC_{3s}.

4.7. Relationship between codon usage bias and compositional constraints in the coding sequences of proto-oncogenes/oncogenes

In order to investigate the relationship between codon usage variation and compositional constraints, we accomplished Pearson's correlation analysis between the values of A, T, G, C, GC and A_3 , T_3 , G_3 , G_3 , G_3 , values, respectively (Table 7).

Chapter 4 **Results**

Significant positive correlation was observed in nucleotide composition, suggesting that nucleotide constraint may influence the codon usage pattern in protooncogenes/oncogenes. In addition, a neutrality plot was drawn (Figure 12) to estimate the extent of directional mutation pressure against selection in the codon usage bias of human proto-oncogenes/oncogenes (Sueoka 1988). In neutrality plot analysis, when a gene is placed on the slope of unity there exists a significant correlation between its GC_{12} (the average value of GC_1 and GC_2) and GC_3 indicating that the gene is under neutral mutation through random selective pressure. But if the gene is under directional mutation pressure, the gene would fall below the slope of unity, i.e. closer to X-axis and further from the Y-axis. Therefore, a regression line with a slope less than 1, indicates that a non neutral mutation pressure affects the codon usage among different genes of the same genome (Necsulea and Lobry 2006, Sueoka and Kawanishi 2000). In our analysis, we observed a significant positive correlation (Pearson r=0.543, p<0.01) between GC_{12} and GC_{3s} which suggested that intragenomic GC mutation bias plays an important role in shaping the codon usage pattern of these genes. The neutrality plot (as shown in figure 12) reveals the results of equilibrium coefficient of mutation and selection. In the plot (Figure 12) some of the points are located in the diagonal distribution and the values of $GC₃$ are in a wide range of distribution, indicating GC_{12} and GC_3 are definitely the affects of mutation bias model (Sueoka 1988). Accordingly, mutation bias might just play a minor role in shaping the codon bias, whereas the natural selection seems to probably dominate the codon bias. Further, we quantify the natural selection and the mutation pressure using regression coefficient. The regression coefficient of GC_{12} to GC_{3} of human proto-oncogene/oncogene is 0.176, indicating the relative neutrality is 17.6 %, while

the relative constraint is 82.4 % for $GC₃$. These results suggest that natural selection played a major role while mutation pressure played a minor role in the codon usage patterns of these genes.

Table 7: Correlation analysis between A, T, G, C, GC contents and A₃, T₃, G₃, C₃, $GC₃$ contents in all the selected coding sequences of proto-oncogenes/oncogenes

	A_3	T_3	G3	C_3	GC ₃
$\overline{\mathsf{A}}$	$r = 0.902**$	$r = 0.876**$	$r = 0.799**$	$r=0.727**$ $r=-0.302*$	
\top	$r = 0.883**$	$r = 0.915**$	$r = 0.766$ **	$r=0.717**$ $r=-0.301*$	
G	$r = 0.638**$	$r = 0.660**$	$r = 0.958**$	$r=0.942**$ $r=0.049$	
C	$r = 0.624***$	$r = 0.632**$	$r = 0.952**$	$r=0.953**$ $r=0.113$	
GC	$r = -0.551**$	$r = -0.508**$	$r = 0.140$	$r = 0.258$	$r = 0.912**$
** $p<0.01$	$*_{p<0.05}$				

Figure 12: Neutrality plot of GC_{12} **versus** GC_3 **. The regression line is** $y = 0.176x +$ 39.48; R^2 = 0.294; GC₁₂: average GC content at first and second codon positions.

4.8. Relationship between codon usage bias and compositional constraints in the coding sequences of tumour suppressor genes

Correlation analysis between the values of A, T, G, C, GC and corresponding A_3 , T₃, G_3 , C_3 , GC_3 values, respectively, were performed in order to investigate the relationship between codon usage variation and compositional constraints (Table 8).

Chapter 4 **Results**

We observed a significant positive correlation between nucleotide compositions, suggesting that nucleotide constraint might influence the codon usage pattern in tumour suppressor genes. Moreover, we performed a neutrality plot analysis of GC_{12} versus $GC₃$ (Figure 13) and in our analysis, we observed a significant positive correlation (Pearson r=0.724, p<0.01) between GC_{12} and GC_3 values which suggested that the intragenomic GC mutation bias might influence the codon usage of these genes. The linear regression coefficient of GC_{12} on GC_{3s} was 0.259 indicating the relative neutrality is 25.9 %, while the relative constraint is 74.1 % for GC3. These results suggest that natural selection played a major role while mutation pressure played a minor role in the codon usage patterns of these genes.

Table 8: Correlation analysis between A, T, G, C, GC contents and A₃, T₃, G₃, C₃, GC₃ contents in all the selected coding sequences of tumor suppressor genes

	A_3	T_3	G3	C_3	GC ₃
A	$r = 0.983**$	$r = 0.982**$	$r = 0.755$ **		$r=0.621**$ $r=-0.410**$
\top	$r = 0.981**$	$r = 0.986**$	$r=0.729**$		$r=0.606**$ $r=-0.431**$
G	$r = 0.826**$	$r = 0.803**$	$r = 0.950**$	$r=0.910**$ $r=-0.097$	
C	$r = 0.743**$	$r = 0.713**$	$r = 0.953**$	r=0.957**	r=0.013
GC	$r = -0.445**$	$r=-0.470**$	$r = 0.144$	$r = 0.308*$	r=0.940**
$*$ p<0.01	$*_{p<0.05}$				

Figure 13: Neutrality plot of GC_{12} **versus** GC_3 **. The regression line is** $y = 0.259x +$ 34.86; R^2 = 0.496; GC₁₂: average GC content at first and second codon positions.

4.9. Effective number of codons (ENC) and its relationship with GC3s values in the coding sequences of proto-oncogenes/oncogenes

The ENC values of coding sequences ranged from 38 to 60 indicating relatively weak codon usage bias among these genes. The $GC₃$ values ranged from 30.4% to 90.8%. The correlation coefficient between ENC and GC_{3s} values showed a significant negative correlation (Pearson $r=-0.835$, $p<0.01$) for each gene, suggesting that genes with higher GC_{3s} values and lower *ENC* values had strong bias. Further, to investigate the relationship between codon usage variations among genes, we plotted the values of ENC versus GC_{3s} (Figure 14) as per Wright (1990). The comparison of actual versus expected distribution in the absence of selection reveals that apart from compositional bias other forces exist which may influence the codon bias. Conversely, if GC_{3s} had been solely responsible for codon usage variation among the genes, the ENC values would have fallen on the ENC -GC $_{3s}$ curve (Wright 1990). The curve line (shown in figure 7) indicates the expected position of genes in which $GC₃$ value is the determining factor in shaping the codon usage pattern. Most

of the genes in our analysis were lying on and close to the reference line, which suggests that GC_{3s} value was the major determining factor of codon usage pattern of these genes. Thus, it is evident that compositional bias might also play a role in defining the codon usage variation in human proto-oncogenes/oncogenes.

4.10. Effective number of codons (ENC) and its relationship with GC3s values in the coding sequences of tumour suppressor genes

The ENC values in the coding sequences of tumour suppressor genes ranged from 35 to 60 indicating relatively weak codon usage bias among these genes. The $GC₃$ values ranged from 25.4% to 94.7%. We also calculated the correlation coefficient between ENC and GC_{3s} values and the result showed a significant negative correlation (Pearson r=-0.695, p<0.01) for each gene. Further, in order to investigate

Chapter 4 **Results**

the relationship between codon usage variations among genes, we plotted the values of ENC versus GC_{3s} (Figure 15) as per Wright (1990).

The comparison of actual versus expected distribution in the absence of selection reveals that apart from compositional bias other forces exist which might influence codon bias. Conversely, if GC_{3s} had been solely responsible for codon usage variation among the genes, the ENC values would have fallen on the ENC -GC $_{3s}$ curve (Wright 1990). The solid curve line (shown in figure 8) indicates the expected position of genes in which GC_3 value is the determining factor in shaping the codon usage pattern. Most of the genes in our analysis were lying on and close to the reference line, representing that GC_{3s} value was the sole determining factor of codon usage pattern of these genes. Thus, it is evident that compositional bias might play a significant role in defining the codon usage variation in human tumour suppressor genes.

Figure 15: Distribution of effective number of codons (ENC) and GC3s in the coding sequences of tumour suppressor genes. The curve indicates the expected curve between ENC and GC contents under random codon usage.

4.11. Frequency of optimal codons for the corresponding amino acids among proto-oncogenes/oncogenes

The frequently used optimal codon (*Fop*) values for the proto-oncogenes/oncogenes were analyzed in order to determine the occurrence of the highest and the lowest frequently used codons for the corresponding amino acid. The average percentage of the Fop value of codons showed that the highest frequently used codons were GCC, AGA, AAC, GAC, TGC, CAG, GAG, GGC, CAC, ATC, CTG, AAG, TTC, CCC, AGC, ACC, TAC and GTG for the amino acids alanine, arginine, asparagine, aspartate, cysteine, glutamine, glutamate, glycine, histidine, isoleucine, leucine, lysine, phenylalanine, proline, serine, threonine, tyrosine and valine respectively (Table 9).

4.12. Frequency of optimal codons for the corresponding amino acids among tumour suppressor genes

To determine the occurrence of the highest and the lowest frequently used codons for each amino acid, the frequently used optimal codon (Fop) values for the tumour suppressor genes were analyzed. The average Fop values of codons showed that the most frequently used codons were GCC, AGA, AAC, GAC, TGC, CAG, GAG, GGC, CAC, ATC, CTG, AAG, TTT, CCT, AGC, ACA, TAC and GTG for the amino acids alanine, arginine, asparagine, aspartate, cysteine, glutamine, glutamate, glycine, histidine, isoleucine, leucine, lysine, phenylalanine, proline, serine, threonine, tyrosine and valine respectively (Table 10).

Table 9: The average of Fop values showing the highest percentage of codon usage for the corresponding amino acid among proto-oncogenes/oncogenes

Table 10: The average of Fop values showing the highest percentage of codon usage for the corresponding amino acid among tumour suppressor genes

4.13. Relative synonymous codon usage in the coding sequences of protooncogenes/oncogenes

The relative synonymous codon usage values of 59 codons in the coding sequences for proto-oncogenes/oncogenes were analyzed excluding the codons ATG (methionine) and TGG (tryptophan). In our calculation, the RSCU value greater than unity indicates the usage of most abundant codon whereas RSCU value less than one reveals the usage of least abundant codon. In addition, RSCU value greater than 1.6 and less than 0.6 indicates the over-represented and the under-represented codon respectively (Wong et al. 2010). The overall RSCU values in the selected coding sequences of proto-oncogenes/oncogenes revealed that 27 codons were most frequently used among the 59 codons and the most predominantly used codons were G/C –ending compared to A/T –ending (Table 11). Besides, it was observed that C –ending codon (15) was mostly favored compared to G –ending codon (7) in the coding sequence of proto-oncogenes/oncogenes. In addition, the most over-represented codon in the coding sequence of proto-oncogenes/oncogenes was CTG encoding leucine amino acid.

4.14. Relative abundance of dinucleotide in the coding sequences of protooncogenes/oncogenes

Literature suggested that dinucleotide bias can influence the overall codon usage patterns in a variety of organisms (Chiusano et al. 2000, Karlin and Burge 1995). The relative abundance of 16 dinucleotides from the coding sequences of eighty-two proto-oncogenes/oncogenes was calculated in order to assess the effect of dinucleotides on the codon usage patterns in these genes under study (Figure 16). The outcome of the analysis showed that CpG dinucleotides were under-represented

70

(mean \pm standard deviation = 0.51 \pm 0.206) whereas GpC dinucleotides were overrepresented ones (mean \pm standard deviation = 1.04 \pm 0.113). Moreover, RSCU values (Table 11) of codons containing the CpG dinucleotide (TCG, CCG, ACG, GCG, CGA, and CGT) except CGC and CGG were the least used codons (RSCU<1.0) for their corresponding amino acid. Similarly, the GpC dinucleotide containing codons (TGC, CGC, GGC, GCC, AGC, GCT) except GCA and GCG were over-represented (RSCU>1.0) in proto-oncogenes/oncogenes under study. Besides, the dinucleotide TpG containing four codons (TGC, CTG, TGC, GTG) and CpA containing four codons (CCA, CAC, CAG, and ACA) were over-represented and most of them were also used as preferred codons for their corresponding amino acid based on RSCU analysis except TTG and TGT for TpG dinucleotide and TCA, CAA, GCA, CAT for CpA dinucleotide in human proto-oncogenes/oncogenes. Previous study reported that the CpA and TpG dinucleotides were over-represented in different organisms. This could be due to the effect of CpG dinucleotide (Bird 1980). Our results also revealed that most of the codons containing CpA and TpG dinucleotides were over-represented in the selected proto-oncogenes/oncogenes under study.

Table 11: Overall relative synonymous codon usage patterns (RSCU) in the coding sequences of proto-oncogenes/oncogenes

a mean values of RSCU based on the synonymous codon usage frequencies of proto-oncogenes/oncogenes; N: Total number of preferred codon; *RSCU>1

Figure 16: Line diagram showing the relative abundance of sixteen dinucleotides in the coding sequence of proto-oncogenes/oncogenes in human

4.15. Trends of codon usage variation among proto-oncogenes/oncogenes

To determine the trends in codon usage variation among human protooncogene/oncogenes, we performed correspondence analysis (COA) based on the RSCU values. In our analysis, we observed that the first principal axis (f_1) accounted for 46.68% of all variations within the gene set, whereas the second axis (f_2) accounted for only 7.86% (Figure 17). The major trends of variation in the entire data can be illustrated in terms of correlation analysis by using primary axis. Therefore, a correlation analysis of the axis 1 (f_1) with the major indices, namely *ENC*, GC, GC3s and CAI values (r= 0.781, -0.933, -0.985 and -0.798, respectively, p<0.01) was done using Spearman's rank correlation method. The above results indicate significant correlation between the primary axes against the four major indices. Hence, the results revealed that nucleotide composition and mutation bias might play major roles in the codon usage of human proto-oncogenes/oncogenes.

Figure 17: Correspondence analysis of RSCU values in human protooncogenes/oncogenes. Each point in the plot represents the distribution of a gene corresponding to the coordinates of the primary and secondary axes of variation.

4.16. Relative synonymous codon usage in the coding sequences of tumour suppressor genes

The relative synonymous codon usage values of 59 codons in the coding sequences for tumour suppressor genes were analyzed excluding the codons ATG (methionine) and TGG (tryptophan). The overall RSCU values in the selected coding sequences of tumour suppressor genes revealed that 32 codons were most frequently used among the 59 codons and the most predominantly used codons were C/T–ending compared to A/G–ending ones (Table 12).

Besides, it was observed that C–ending codon (12) was mostly favored compared to T –ending codon (8) followed by G–ending (7) and A–ending codon (5) in the coding sequences of tumour suppressor genes. The most over-represented codon (highest RSCU value) was CTG encoding the amino acid leucine.

4.17. Relative abundance of dinucleotide in the coding sequences of tumour suppressor genes

The relative abundance of 16 dinucleotides from the coding sequences of sixty three tumour suppressor genes were calculated in order to assess the effect of dinucleotides on the codon usage patterns in these genes under study (Figure 18). The results of the analysis showed that CpG dinucleotides were under-represented (mean \pm standard deviation = 0.49 \pm 0.210) whereas GpC dinucleotides were overrepresented (mean \pm standard deviation = 1.20 \pm 0.111). Moreover, RSCU values (Table 12) of codons containing the CpG dinucleotide (TCG, CCG, ACG, GCG, CGA, CGC, CGG and CGT) were the least used codons further (RSCU<1.0) for their corresponding amino acid. Similarly, the GpC dinucleotide containing codons (TGC, GCA, GGC, GCC, AGC, GCT) except CGC and GCG were over-represented (RSCU>1.0) in tumour suppressor genes under study. In addition, the dinucleotide TpG containing four codons (TGC, CTG, TGC, GTG) and CpA containing four codons (CCA, CAC, CAG, GCA and ACA) were over-represented and most of them were also used as preferred codons for their corresponding amino acid based on RSCU analysis except TTG and TGT for TpG dinucleotide and TCA,CAA, CAT for CpA dinucleotide in tumour suppressor genes. The over-representation of CpA and TpG dinucleotides was also reported earlier in different organisms (Bird 1980).

Table 12: Overall relative synonymous codon usage patterns (RSCU) in the coding

sequences of tumour suppressor genes

amean values of RSCU based on the synonymous codon usage frequencies of tumour suppressor genes; N: Total number of preferred codon; *RSCU>1

Figure 18: Line diagram showing the relative abundance of sixteen dinucleotides in the coding sequence of tumour suppressor genes in human

4.18. Trends of codon usage variation among tumour suppressor genes

Correspondence analysis (COA) based on RSCU values was performed in order to determine the trends in codon usage variation among human tumour suppressor genes. We observed that the first principal axis (f_1) accounted for 56.10% of all variations within the gene set, whereas the second axis (f_2) accounted for only 5.17% variation (Figure 19). The major trends of variation in the entire data can be illustrated in terms of correlation analysis by using primary axis. Therefore, a correlation analysis of the axis 1 (f_1) with the major codon bias indices, namely *ENC*, GC, GC3s and CAI values (r= -0.385, 0.947, 0.982 and 0.837, respectively, p<0.01) was done using Spearman's rank correlation method. The above results indicate significant correlation between the primary axis and the four major indices. Hence, the results revealed that both nucleotide composition and mutation bias might play major roles in the codon usage of human tumour suppressor genes.

Figure 19: Correspondence analysis of RSCU values in human tumour suppressor genes. Each point in the plot represents the distribution of a gene corresponding to the coordinates of the primary and secondary axes of variation.

4.19. Prediction of proto-oncogenes/oncogenes expression level and codon usage bias

Codon adaptation index (CAI) is a quantitative measure used for the prediction of gene expression on the basis of extent of bias towards codon sequence (Behura and Severson 2012, Gupta et al. 2004). In our analysis, CAI values ranged from 0.715 to 0.887 (Figure 20) with a mean value of 0.801 and a standard deviation of 0.039. The magnitude of CAI values indicates that most of the genes selected in the present study are highly expressive in cell.

Moreover, a significant positive correlation was observed between CAI and GC_{3s} $(r=0.847, P<0.01)$ and between *CAI* and GC $(r=0.715, P<0.01)$ content values. Furthermore, significant negative correlation (r=-0.627, P<0.01) also observed between ENC and CAI values (Figure 21).

Figure 20: Trends of the CAI values in the coding sequences of protooncogenes/oncogenes

Figure 21: Correlation analysis between CAI values and ENC, GC, GC3 values. [A] Correlation between CAI and ENC. [B] Correlation between CAI and GC contents. [C] Correlation between CAI and $GC₃$ values.

4.20. Relationship between codon usage and CAI values of protooncogenes/oncogenes

To investigate the relationship between the codon usage variation and the level of gene expression among the selected coding sequences of human protooncogenes/oncogenes, the correlation coefficient between codon usage and CAI was estimated using heat map (Figure 22). The results showed that almost all G/C – ending codons are positively correlated with CAI and vice versa for A/T –ending codons. This indicates that gene expression increases with the increase in usage of G/C –ending codons. However, two G–ending codons namely TTG (leucine) and CAG (glutamine) showed negative correlation between codon usage and gene expression as measured by CAI.

Figure 22: Heat maps of correlation coefficient of codons with CAI. The color coding red represents the positive correlation, green as negative correlation. The black fields are stop codons (TAA, TAG, TGA) and non-degenerate codons (ATG, TGG) in the coding sequences of the eighty-two proto-oncogenes/oncogenes under study.

4.21. Prediction of translational efficiency as measured by tAI and its relationship with codon usage in proto-oncogenes/oncogenes pool

The analysis of tAI for the coding sequences in a genome acquires importance only when the translational selection process plays a major role in the shaping of the codon usage of the particular genome (Man et al. 2006). We estimated tAI values for human proto-oncogenes/oncogenes pool (Jung and McDonald 2011) which ranged from 0.326 to 0.414 with a mean of 0.360. We accomplished a correlation analysis between codon usage and tAI and represented the same in a heat map (Figure 23A). The results showed that nearly all codons ending with G/C base were positively correlated with tAI and vice versa for all A/T ending codons. Besides, we observed a significant negative correlation (r=-0.545, p<0.01) between tAI and effective number of codons (ENC) and significant positive correlation between tAI and GC3s, an indicator of the extent of base composition bias $(r=0.382, p<0.01)$ (Figure 23 -B & C). In addition, a significant positive correlation (Pearson, r=0.676, p<0.01) was observed between the two parameters tAI and CAI of the coding sequences which indicated that the expression of proto-oncogenes/oncogenes, was significantly influenced by the genomic tRNA pool.

81

Figure 23. Relationship between tAI and codon usage bias in protooncogene/oncogene pool. [A] Correlation coefficient between codon usage and tAI. The rectangular red color box indicates the positive correlation, green as negative correlation; black boxes are non-degenerate codons (ATG, TGG) and three termination codons (TAA, TAG, TGA). [B] Correlation coefficient between tAI and ENC. [C] Correlation coefficient between tAI and GC3s.

4.22. Prediction of tumour suppressor gene expression level and codon bias

The expression of tumour suppressor gene was measured using CAI values which ranged from 0.713 to 0.858 (Figure 24) with a mean of 0.775 and a standard deviation of 0.037. The CAI values indicate that most of the tumour suppressor genes selected for the present study are highly expressed in cells.

In addition, we observed significant negative correlation between CAI and ENC (r=-0.515, P<0.01) values, significant positive correlation between CAI and $GC₃$ (r=0.839, P<0.01) as well as between CAI and GC contents (r=0.730, P<0.01) in tumour suppressor genes (Figure 25).

Figure 24: Trends of the *CAI* values in the coding sequences of tumour suppressor genes

Figure 25: Correlation analysis between CAI values and ENC, GC, GC3 values. [A] Correlation between CAI and ENC. [B] Correlation between CAI and GC contents. [C] Correlation between CAI and $GC₃$ values.

4.23. Relationship between codon usage and CAI values of tumour suppressor genes

To investigate the relationship between the codon usage variation and gene expression level among the selected coding sequences of human protooncogenes/oncogenes, the correlation coefficient between codon usage and CAI was analyzed using heat map (Figure 26). The result showed that almost all the G/C –ending codons are positively correlated with CAI and vice versa for A/T–ending codons. This suggests that gene expression increases with the increase in usage of G/C–ending codons. However, two G–ending codons namely AGG (arginine) and TTG (leucine) showed negative correlation between codon usage and gene expression as measured by CAI.

Figure 26: Heat maps of correlation coefficient of codons with CAI. The color coding red represents the positive correlation, green as negative correlation. The black fields are stop codons (TAA, TAG, TGA) and non-degenerate codons (ATG, TGG) in the coding sequences of the sixty-three tumour suppressor genes under study.

4.24. Prediction of translational efficiency as measured by tAI and its relationship with codon usage in tumour suppressor gene pool

We calculated tAI values for human tumour suppressor gene pool (Jung and McDonald 2011) which ranged from 0.382 to 0.402 with a mean value of 0.356. We performed a correlation analysis between codon usage and tAI and represented it in a heat map (Figure 27 A). The results showed that nearly all codons ending with G/C base were positively correlated with tAI and vice versa for all A/T ending codons.

Besides, we observed a significant positive correlation (r=-0.382, p<0.01) between tAI and effective number of codons (ENC) as well as between tAI and GC3s, an indicator of the extent of base composition bias (r=0.511, p<0.01) (Figure 27-B & C). Moreover, we observed a significant positive correlation (r=0.625, p<0.01) between the two parameters tAI and CAI for coding sequences. This revealed that the expression of tumour suppressor genes (CAI) was significantly influenced by the genomic tRNA pool.

Figure 27. Relationship between tAI and codon usage bias in tumour suppressor gene pool. [A] Correlation coefficient between codon usage and tAI. The rectangular red color box indicates the positive correlation, green as negative correlation; black boxes are non-degenerate codons (ATG, TGG) and three termination codons (TAA, TAG, TGA). [B] Correlation coefficient between tAI and ENC. [C] Correlation coefficient between tAI and GC3s.